Viscosic acid (5). Oil (8 mg). UV $\lambda_{\text{max}}^{\text{McOH}}$ nm: 215 (log ε 4.3); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 2950, 2840, 2600 (br sh), 1695, 1620, 1445, 1380, 1100, 1050, 980, 820, 750; ¹H NMR (400 MHz, CDCl₃) see text; MS (probe) 70 eV (rel. int.): 264 [M]⁺ (C₁₆H₂₄O₃) (methyl ether of viscosic acid) (18), 250 [M - CH₂]⁺ (16), 249 [M - Me]⁺ (68), 233 (13), 203 (20), 156 (31), 117 (98).

Epoxidation of costic acid methyl ester. p-Chloroperbenzoic acid (10 mg) was added to costic acid methyl ester (5 mg) in 1 ml CH₂Cl₂ in the presence of NaHCO₃ (10 mg). The mixture was left at room temp. for 1 hr. The ¹H NMR spectrum of the resulting compound was identical with that of compound 5.

Acknowledgements—This work was supported by the Health Sciences Institute of the University of Istanbul (Grant No. 114). The authors thank Dr. J. Jakupovic (Berlin) for his kind help in structure determination.

REFERENCES

- Baytop, T (1984) Therapy with Medicinal Plants in Turkey, p. 167. Sanal Press, Istanbul.
- 2. Gad, S. and Yoel, K. (1970) J. Med. Chem. 13, 1221.
- Suslugas, C., Balansard, G., Julien, J., Gasquet, M., Timon-Davis, P. and Ross, J. C. (1980) Herba Hung. 19, 19; Chem. Abstr. 94, 20290.
- Herz, W., Hiroaki, C. and Tether, L. H. (1966) J. Org. Chem. 31, 1632.
- Bohlmann, F., Czerson, H. and Schöneweiss, S. (1977) Phytochemistry 11, 2859.
- Zdero, C., Bohlmann, F., King, R. M. and Robinson, H. (1987) Phytochemistry 26, 1207.
- 7. Öksüz, S. (1976) Planta Med. 343.
- 8. Grande, M., Piera, F., Cuenca, A., Torres, P. and Bellido, I. S. (1985) Planta Med. 414.

Phytochemistry, Vol. 26, No. 4, pp. 1224-1226, 1987. Printed in Great Britain. 0031-9422/87 \$3.00 + 0.00 Pergamon Journals Ltd.

A FLAVONOL GLYCOSIDE FROM LYSIMACHIA MAURITIANA

KEN YASUKAWA and MICHIO TAKIDO

Department of Pharmacy, College of Science and Technology, Nihon University, Tokyo 101, Japan

(Revised received 29 September 1986)

Key Word Index—*Lysimachia mauritiana*; Primulaceae; mauritianin; kaempferol-3-*O*-(2,6-di-*O*-α-rhamno-pyranosyl-β-galactopyranoside); flavonol glycoside.

Abstract—From the whole plant of Lysimachia mauritiana, a new flavonol glycoside (mauritianin) was isolated together with hyperin, kaempferol-3-O-robinobioside and kaempferol-3-O- α -rhamnopyranosyl-(1-2)- β -galactopyranoside. The structure of mauritianin was established as kaempferol-3-O-(2,6-di-O- α -rhamnopyranosyl- β -galactopyranoside).

INTRODUCTION

Flavonol glycosides of the kaempferol, quercetin and myricetin type have already been isolated from the genus Lysimachia, a member of the Primulaceae (L. vulgaris [1, 2], L. punctata [3] and L. nummularia [4]). Initial chemical investigations of L. mauritiana led to the isolation and structure elucidation of sapogenins [5, 6]. In this paper we report the isolation and characterization of four flavonol glycosides from this plant. Hyperin and kaempferol-3-O-rhamnosyl galactosides are reported for the first time in Lysimachia; mauritianin is a new compound, which an anti-tumour promoter [7].

RESULTS AND DISCUSSION

The concentrated methanol extract prepared from the air dried plant was extracted successively with ethyl acetate and n-butanol. One flavonol glycoside (compound 1) was isolated on a Sephadex LH-20 column using water-methanol from the ethyl acetate fraction, and another three flavonol glycosides (compounds 2-4) were isolated by the same method from the n-butanol fraction.

Compound 1, yellow needles, mp 238.5-239°, gave positive ferric chloride and Mg + HCl tests and was identified as hyperin by direct comparison with an authentic sample.

Short Reports 1225

Compound 2, pale yellow needles, mp 198-200°, also gave positive ferric chloride and Mg + HCl tests. The acid hydrolysis products of 2 showed the presence of kaempferol and the two sugars, galactose and rhamnose, by TLC. 2 was subsequently identified as kaempferol-3-Orobinobioside by direct comparison with an authentic sample.

The similarity in the chromophore of compound 3 to that of compound 2 was established by comparison of their colour reactions, acid hydrolysis products and UV spectrum. ¹³C NMR also showed that 3 had galactose and rhamnose units in its structure, the only significant difference from 1 being an upfield shift of 4.1 ppm for the C-2" of galactose and a downfield shift of 3.5 ppm for the C-1" of galactose (Table 1). These shifts are analogous to those reported [8] when the C-2" of galactose is rhamnosylated in a flavonol glycoside. The structure of compound 3 was therefore determined as kaempferol-3-O-α-rhamnopyranosyl-(1-2)-β-galactopyranoside.

Compound 4, pale yellow needles, mp 202-204°, again gave positive ferric chloride and Mg + HCl tests. The acid hydrolysis products of 4 showed the presence of kaempferol, galactose and rhamnose on TLC. Kaempferol was identified by direct comparison with an authentic sample. The ¹³C NMR of 4 in DMSO-d₆ also confirmed that it was a glycoside of kaempferol. The 13C NMR shifts of the aglycone part of 4 corresponded well to the shifts for kaempferol, the only significant difference being an upfield shift of 2.4 ppm for the C-3. This shift is analogous to that reported [8] when the 3-hydroxy group is glycosylated in a flavonol glycoside. UV shifts on the addition of sodium acetate and aluminium chloride also showed the position of linkage between sugar and aglycone as C-3. The ¹³C NMR of 4 is similar to that of compounds 1-3, the only significant difference being an upfield shift of 4.1 and 5.4 ppm, respectively, for the C-2" and C-6" of galactose because it is a rhamnosylated flavonol glycoside [9]. Thus, the structure of compound 4 (named mauritianin) was determined as kaempferol-3-O- $(2,6-di-O-\alpha-rhamnopyranosyl-\beta-galactopyranoside)$. The same trisaccharide has already been found in another kaempferol glycoside isolated from Rhamnus nakaharai but in this case the sugar is attached to the 4'-OH [9].

EXPERIMENTAL

All mps are uncorr. 1 H and 13 C NMR spectra were recorded at 100 and 25.5 MHz, respectively, chemical shifts are given in δ (ppm) with TMS as int. std. CC was carried out using Sephadex LH-20 (Pharmacia). TLC on Avicel SF (Funakoshi) was performed with n-BuOH-HOAc-H₂O (3:1:1).

Plant material. L. mauritiana Lam. was collected at Miura beach, Kanagawa, Japan in the autumn of 1983.

Extraction and isolation. Dried whole plant (1.5 kg) was extracted with MeOH and coned to a dark residue, which was then macerated with hot H_2O and filtered. The H_2O soln was extracted with EtOAc followed by n-BuOH. The EtOAc extract (2 g) was then subjected to CC on Sephadex LH-20, using H_2O -MeOH as eluant, to yield compound 1 (10 mg). The n-BuOH extract (10 g) was analysed by the same method, to yield compounds 2 (10 mg), 3 (20 mg) and 4 (102 mg).

Hyperin. Recrystallization (H₂O-MeCN) gave yellow needles, mp 238.5–239°. Analysis: calcd: $C_{21}H_{20}O_{12} \cdot H_2O$: C, 52.33; H, 4.76; found: C, 52.29; H, 4.60. Dark green colour after FeCl₃ reaction, pale red colour in Mg + HCl test. UV $\lambda_{\rm min}^{\rm EIOH}$ nm: 258, 270 sh, 293sh, 364. IR $\nu_{\rm min}^{\rm KB}$ cm $^{-1}$: 3300 (OH), 2900 (C-H), 1655 (C=O), 1605 (C=C), 1060 (C-O). ¹H NMR (DMSO-d₆) δ ppm: 12.65 (1H, s, OH-5), 7.60 (2H, dd, J = 2.0 Hz, J = 8.3 Hz, H-6'), 7.59 (1H, d, J = 2.0 Hz, H-2'), 6.86 (1H, d, J = 8.3 Hz, H-6'), 6.42 (1H, d, J = 1.9 Hz, H-8), 6.21 (1H, d, J = 1.9 Hz, H-6), 5.52 (1H, d, J = 7.3 Hz, galactosyl H-1). ¹³C NMR (DMSO-d₆, 90°) δ ppm: 177.6 (C-4), 164.2 (C-7), 161.3 (C-5), 156.5 (C-2, C-9), 148.4 (C-4'), 144.8 (C-3'), 133.8 (C-3), 98.8 (C-6), 93.6 (C-8). For chemical shifts of sugar units see Table 1.

Kaempferol-3-O-robinobioside. Recrystallization (H₂O-MeCN) gave pale yellow needles, mp 198–200°. Analysis: calcd: $C_{27}H_{30}O_{16} \cdot 3/2H_2O$: C,50.80; H,5.17; found; C,50.87; H,5.22. Dark green colour after FeCl₃ reaction, pink colour in Mg + HCl test. UV λ_{\max}^{EIOH} nm: 263, 301sh, 349. IR ν_{\max}^{KB} cm⁻¹: 3400 (OH), 2950 (C-H), 1660 (C=O), 1605 (C=C), 1060 (C-O). ¹H NMR (DMSO-d₆) δ ppm: 12.58 (1H, s, OH-5), 8.00 (2H, d, J = 8.3 Hz, H-2', H-6'), 6.89 (2H, d, J = 8.3 Hz, H-3', H-5'), 6.40 (1H, d, J = 1.9 Hz, H-8), 6.23 (1H, d, J = 1.9 Hz, H-6), 5.56 (1H, d, J = 7.3 Hz, galactosyl H-1), 5.05 (1H, s, rhamnosyl H-1), 1.05 (3H, d, J = 5.4 Hz, rhamnosyl Me). ¹³C NMR (DMSO-d₆, 90°) δ

Table 1. 13C NMR spectral data of sugar units of compounds 1-4 (25.5 MH	łz,
DMSO-d ₆ , TMS as internal standard)	

Galactose C-1"	102.6	102.2	99.1	4	
				99.2	
C-2"	71.5	71.5	75.6	75.6	_
C-3"	73.6	73.4	74.1	73.7	_
C-4"	68.2	68.4	68.2	68.3	
C-5"	75.9	73.9	75.9	74.0	
C-6"	60.4	65.9	60.4	65.8	
Rhamnose C-1		100.4	100.7	100.7	100.3
C-2	_	71.0	70.9	70.9	70.9
C-3	_	70.3	70.8	70.8	70.5
C-4	_	72.6	72.3	72.3	72.3
C-5	_	68.4	68.7	68.8	68.3
C-6	_	17.6	17.2	17.2	17.7

1226 Short Reports

ppm: 177.5 (C-4), 164.3 (C-7), 161.3 (C-5), 160.0 (C-4'), 156.6 (C-2, C-9), 133.8 (C-3), 130.9 (C-2', C-6'), 121.1 (C-1'), 115.4 (C-3', C-5'), 104.3 (C-10), 99.0 (C-6), 93.9 (C-8). Chemical shifts of sugar units are given in Table 1.

Hydrolysis of 2. Compound 2 (3 mg) treated with 2 N HCl at 100° for 3 hr gave kaempferol, galactose and rhamnose. Kaempferol was identified by direct comparison with an authentic sample. The sugars were identified by TLC with authentic samples. $R_f = 0.44$ and 0.30 (rhamnose and galactose $R_f = 0.44$ and 0.30).

Kaempferol-3-O- α -rhamnopyranosyl-(1-2)- β -galactopyranoside. Recrystallization (H2O-MeCN) gave pale yellow needles, mp 202-204°. Analysis: calcd: C27H30O16·9/5H2O: C, 50.73; H, 5.57; found: C, 50.44; H, 5.27. Dark green colour with FeCl₃ reaction, pink colour in Mg+HCl test. UV \(\lambda \) EtOH nm: 262, 300sh, 350. λ EtOH + AlCl₃ nm: 272, 300, 320sh, 345, 395. λ EiOH + NaOAc nm: 263, 300sh, 349. IR ν KBr cm⁻¹: 3400 (OH), 2950 (C-O), 1660 (C=O), 1610 (C=C), 1060 (C-O). 1H NMR (DMSO- d_6) δ ppm: 12.55 (1H, s, OH-5), 8.07 (2H, d, J = 8.3 Hz, H-2', C-6'), 6.85 (2H, d, J = 8.3 Hz, H-3', H-5'), 6.39 (1H, d, J= 1.9 Hz, H-8), 6.17 (1H, d, J = 1.9 Hz, H-6), 5.65 (1H, d, J= 7.3 Hz, galactosyl H-1), 5.04 (1H, s, rhamnosyl H-1), 0.73 (3H, d, J = 5.4 Hz, rhamnosyl Me). ¹³C NMR (DMSO- d_6 , 90°) δ ppm: 177.5 (C-4), 164.8 (C-7), 159.8 (C-4'), 156.5 (C-2), 156.0 (C-9), 132.9 (C-3), 130.6 (C-2', C-6'), 121.2 (C-1'), 115.1 (C-3', C-5'), 104.0 (C-10), 98.9 (C-6), 93.7 (C-8). Chemical shifts of sugar units are given in Table 1.

Hydrolysis of 3. Compound 3 (5 mg) treated with 2 N HCl at 100° for 3 hr gave kaempferol, galactose and rhamnose. Kaempferol was identified by direct comparison with an authentic sample. The sugars were identified as rhamnose and galactose by TLC.

Kaempferol-3-O-(2,6-di-O-α-rhamnopyranosyl-β-galactopyranoside) (mauritianin). Recrystallization (H₂O-MeCN) gave pale yellow needles, mp 202–204°. Analysis: calcd: $C_{33}H_{44}O_{19} \cdot 9/2H_2O$: C,48.14; H,5.71; found: C,48.23; H,6.01. Dark green colour with FeCl₃ reaction, pink colour in Mg + HCl test. UV λ_{\max}^{EiOH} nm: 264, 300sh, 348. $\lambda_{\max}^{EiOH} + \lambda_{\max}^{ICOH} +$

¹H NMR (DMSO- d_o) δppm: 12.70 (1H, s, OH-5), 8.05 (2H, d, J = 8.3 Hz, H-2', H-6'), 6.88 (2H, d, J = 8.3 Hz, H-3', H-5'), 6.41 (1H, d, J = 1.9 Hz, H-6), 6.21 (1H, d, J = 1.9 Hz, H-8), 5.56 (1H, d, J = 7.3 Hz, galactosyl H-1), 5.05 (2H, s, rhamnosyl H-1, H-1'), 1.05 (3H, d, J = 5.4 Hz, rhamnosyl Me), 0.79 (3H, d, J = 5.4 Hz, rhamnosyl Me'). ¹³C NMR (DMSO- d_o , 90°) δppm: 177.5 (C-4), 164.0 (C-7), 161.3 (C-5), 159.7 (C-4'), 156.5 (C-2, C-9), 132.9 (C-3), 130.6 (C-2', C-6'), 121.3 (C-1'), 115.2 (C-3', C-5'), 104.2 (C-10), 98.9 (C-6), 93.8 (C-8). Chemical shifts of the sugar units are given in Table 1.

Hydrolysis of 4. Compound 4 (5 mg) treated with 2 N HCl at 100° for 3 hr gave an aglycone and sugars. The aglycone was identified as kaempferol by direct comparison with an authentic sample. The sugars were identified as rhamnose and galactose by TLC.

Acknowledgements—We are grateful to Professor T. J. Mabry, Department of Botany, University of Texas at Austin, for a generous gift of kaempferol-3-O-robinobioside. We thank Y. Kimura, Department of Pharmacy, Nihon University, for IR and Dr. T. Takido, Analytical Centre, College of Science and Technology, Nihon University, for NMR spectra.

REFERENCES

- Rzadkowska-Bodalska, H. and Olechnowicz-Stepien, W. (1975) Pol. J. Pharmacol. Pharm. 27, 345.
- Prum, N., Pichon, P. and Raynaud, J. (1972) Plant. Med. Phytother. 6, 267.
- 3. Mendez, J. (1970) Experientia 26, 108.
- 4. Popov, V. I. (1967) Tr. Leningrad. Khim.-Farm. Inst. 21, 221.
- 5. Usmanghani, K. (1977) Pak. J. Sci. Ind. Res. 20, 393.
- Kitagawa, I., Matsuda, A. and Yosioka, I. (1972) Chem. Pharm. Bull. 20, 2226.
- Yasukawa, K., Takeuchi, M., Sato, Y., Nitta, K. and Takido, M. (1985) Proc. Jpn. Cancer Assoc., 44th Meet. 66.
- Markham, K. R., Ternai, B., Stanley, R., Geiger, H. and Mabry, T. J. (1978) Tetrahedron 34, 1389.
- Lin, C. N., Arisawa, M., Shimizu, M. and Morita, N. (1982) Phytochemistry 21, 1466.